

UC Davis

UC Davis Previously Published Works

Title

Report on the National Eye Institute's Audacious Goals Initiative: Creating a Cellular Environment for Neuroregeneration.

Permalink

<https://escholarship.org/uc/item/9613v6nq>

Journal

eNeuro, 5(2)

ISSN

2373-2822

Authors

Burns, Marie E

Stevens, Beth

Publication Date

2018-03-01

DOI

10.1523/eneuro.0035-18.2018

Peer reviewed

Report on the National Eye Institute's Audacious Goals Initiative: Creating a Cellular Environment for Neuroregeneration

 Marie E. Burns,¹ and Beth Stevens,^{2,3} for the Audacious Goals Initiative Workshop Panelists

DOI:<http://dx.doi.org/10.1523/ENEURO.0035-18.2018>

¹Center for Neuroscience and Departments of Ophthalmology and Vision Science and Cell Biology and Human Anatomy, University of California Davis, Davis, CA 95616, ²F.M. Kirby Neurobiology Center, Boston Children's Hospital and Harvard Medical School, Boston, MA, and ³Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, MA 02115

Abstract

The cellular environment of the CNS is non-permissive for growth and regeneration. In the retina, transplantation of stem cells has been limited by inefficient survival and integration into existing circuits. In November 2016, as part of the National Eye Institute's Audacious Goals Initiative (AGI), a diverse collection of investigators gathered for a workshop devoted to articulating the gaps in knowledge, barriers to progress, and ideas for new approaches to understanding cellular environments within the retina and how these environments may be manipulated. In doing so, the group identified the areas of (1) retinal and optic nerve glia, (2) microglia and inflammation, and the (3) extracellular matrix (ECM) and retinal vasculature as key to advancing our understanding and manipulation of the retinal microenvironments. We summarize here the findings of the workshop for the broader scientific community.

Key words: eye; glia; inflammation; microglia; regeneration; retina

Introduction

The era of cell transplantation and neuroregeneration is today becoming more science than science fiction. The formation of laminated retinal organoids in culture (Meyer et al., 2009, 2011; Eiraku et al., 2011; Zhong et al., 2014; Shirai et al., 2016; Wiley et al., 2016; Reichman et al., 2017; Wahlin et al., 2017) and stem cell transplantation studies are increasingly commonplace. However, the next vital step is to integrate these cells efficiently into a neural

circuit or microenvironment supporting a circuit that is otherwise losing function. A way to control the cellular environment to promote transplanted cell survival, differentiation, and integration into the surviving retinal circuitry may be most important for the success of regenerative medicine. To tackle the question of how to create a cellular environment for neuroregeneration, workshop participants were charged to consider (1) the role of astrocytes and microglia as positive or negative mediators in regeneration and synapse formation, (2) the importance of vasculature and extracellular matrix (ECM) in regeneration, and (3) the use of scaffolds and other extrinsic tools to support regeneration and reconnection of retinal neurons.

To begin to address these issues, it is first important to recognize some of the unique features of the retina and its place in the posterior eye that present physical barriers and create distinct areas within the neural tissue that define unique environmental subcompartments (Fig. 1), especially with regard to lamination of the retinal tissue. For example, the distal-most, or outermost layer of the retina is comprised of rod and cone photoreceptors, whose survival and disk morphogenesis depends intimately on the overlying retinal pigment epithelial (RPE) cells, the retinal choroid plexus, and beyond these, a very important component of the blood-retinal barrier (BRB),

Received January 21, 2018; accepted March 1, 2018; First published April 6, 2018.

The authors report no conflict of interest.

Author contributions: M.E.B. and B.S. wrote the paper, summarizing the insights of the AGI Workshop Panelists.

The workshop was hosted by the National Eye Institute.

Acknowledgments: We thank workshop panelists for lively cross-disciplinary discussions and manuscript contributions; readers are directed to <https://nei.nih.gov/audacious/creating-cellular-environment-neuroregeneration> for a full list of participants. We also thank Kaitryn Ronning (University of California Davis) for help with Figure 1.

Correspondence should be addressed to Marie E. Burns at the above address, E-mail: meburns@ucdavis.edu.

DOI:<http://dx.doi.org/10.1523/ENEURO.0035-18.2018>

Copyright © 2018 Burns and Stevens

This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution and reproduction in any medium provided that the original work is properly attributed.

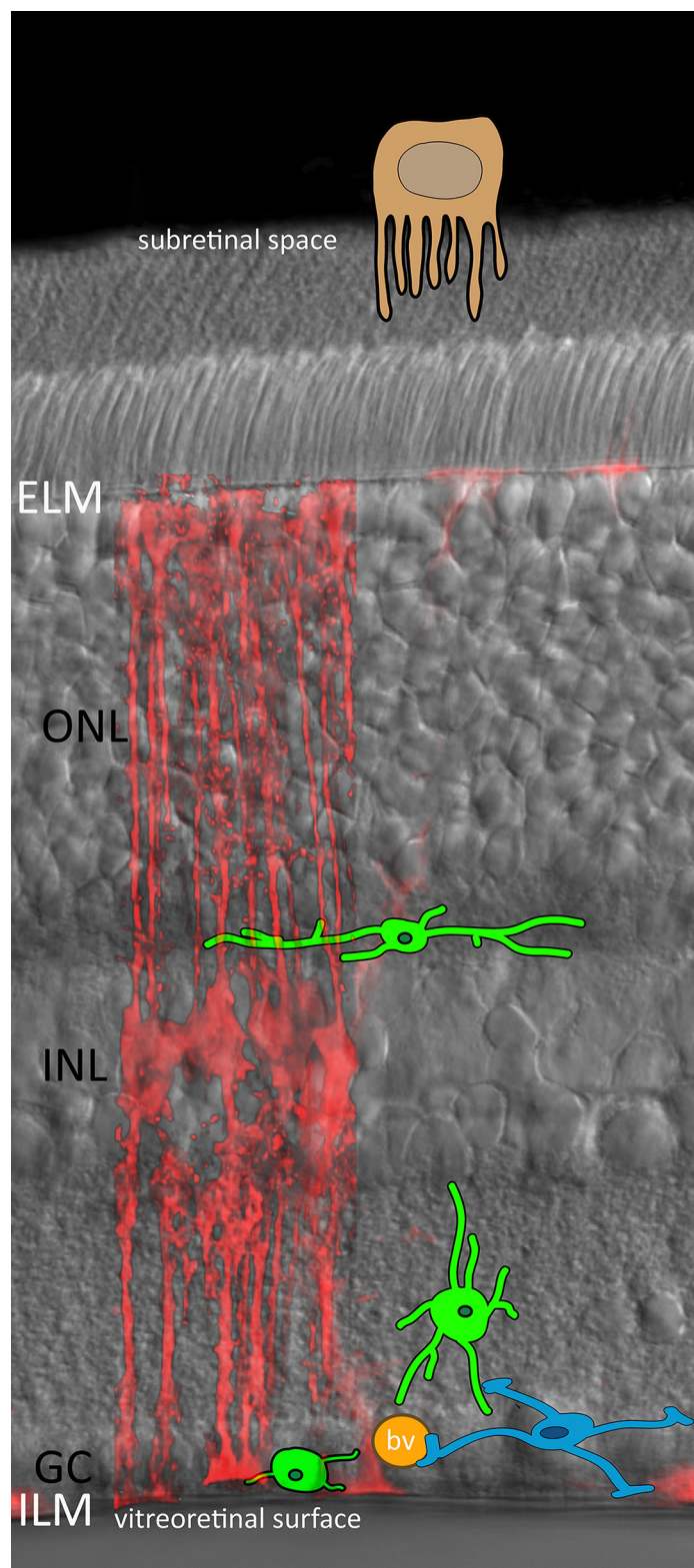


Figure 1. Schematic of local microenvironments within the retina, based on a fixed vibratome section of a mouse retina expressing a viral construct (Shh10-GFAP-mCherry) driving mCherry expression in Müller glial cells (red). Microglia (green) are localized in the synaptic layers between the outer nuclear layer (ONL) and inner nuclear layer (INL) as well as the ganglion cell layer (GC). The subretinal space is subtended by RPE cells (beige) and the ELM secreted by the Müller cells. The Müller cells also secrete the matrix constituting the ILM, which serves as a barrier at the vitreoretinal surface. Astrocytes (blue) are abundant in the nerve fiber layer, where they interact extensively with GCs and blood vessels (bv, orange).

Bruch's membrane. The proximal retina likewise has a clear anatomical barrier, the internal limiting membrane (ILM), which is composed of collagen and other ECM components secreted by the Müller cell endfeet, which keep the ionic and protein milieu of the CNS distinct from that of vitreous, a boundary known as the vitreo-retinal surface. Further, the optic nerve head, where unmyelinated ganglion cell axons exit the eye in forming the nerve proper, also presents challenges to regeneration. These include age-related changes in the inflammatory microenvironment, especially with regard to microglia, astrocytes, bioenergetic capacity, and oxidative stress.

Within this context, we will first discuss the state of the science regarding retinal glia (Müller cells and astrocytes), the role of resident immune cells (microglia) and neuroinflammation, and the functions of the ECM and BRB. Then we will consider the current best strategies and greatest barriers for progress in these areas. It was clear from the workshop that an improved understanding of how these cells and niches maintain the retinal environment, and how injury and disease perturbs those functions, will enable more successful transplantation and neuroregeneration. Thus, this report adds an additional perspective to those of the previous four Audacious Goals Initiative (AGI) workshops and highlights areas for researchers to address for manipulating the cellular environment so that regenerative strategies have maximal effectiveness.

Results

Retinal glia

Glial cells are classically considered the support cells of the nervous system because of their essential role in providing nutrients and growth factors, structural support, maintenance and regulation of the blood-brain and BRB and tissue homeostasis. Glia are particularly adept at sensing and responding to microenvironmental changes. For example, glial expression of aquaporins and TRP channels (Jo et al., 2015) transduce mechanical stress into intracellular calcium signaling and ECM remodeling (Ryskamp et al., 2014; Choi et al., 2015). In recent years, glia have also emerged as active participants capable of refining circuit properties through gliotransmission, promotion of synaptogenesis, synaptic plasticity, and modulation of blood flow through vasoconstriction/dilation in response to changes in electrical activity (Newman, 2015; Giannoni et al., 2018). Elsewhere in the nervous system they also sustain myelination (oligodendrocytes and Schwann cells) and in some instances prevent or impede axon regeneration, either through pro-inflammatory signaling or formation of glial scars (Adams and Gallo, 2018).

The two most abundant glial cell types of the retina, Müller cells and astrocytes, have distinct morphological and physiological characteristics. Müller cells uniquely span all of the retinal layers, from the ILM created by their endfeet at the vitreoretinal surface to the adhesion junctions they make with each other just distal to the outer nuclear layer, called the outer limiting membrane. The distinct, specialized microenvironments of the retina and the subretinal space are unique in their ECM and ionic composition, which the Müller cells help to maintain. Müller

cells shuttle potassium ions from the photoreceptors at outer limiting membrane to the vitreoretinal surface, and are essential for maintaining the accompanying changes in osmotic balance (for review, see Sarthy and Ripps, 2001; Bringmann et al., 2006). The intimate contacts between Müller cells and cone photoreceptors also permits retinoid exchange that helps to maintain visual pigment and cone signaling in bright light (Xue et al., 2015). Unlike glia elsewhere in the brain, Müller cells are deficient in pyruvate kinase and rely on metabolites from photoreceptors and other retinal neurons to fuel their mitochondria, one of their many metabolic oddities needed to sustain the demanding outer retinal environment (for review, see Hurley et al., 2015). Müller glia may hold the secret to unlocking neuroregenerative capacity in the retina since they themselves can divide and produce neural progenitors throughout adulthood in fish (Lenkowski and Raymond, 2014) and after injury in chicken (Fischer and Bongini, 2010). This neurogenic state has recently been successfully induced in mice through the ectopic expression in Müller cells of the proneural transcription factor *Ascl1* (Jorstad et al., 2017).

Retinal astrocytes are found only in the ganglion cell and nerve fiber layers (Büssow, 1980), where they help to control neural activity, contribute to the integrity of the BRB, and regulate vascular tone (for review, see MacVicar and Newman, 2015). Astrocytes form close contacts with the unmyelinated segments of retinal ganglion cell (RGC) axons, where they may play a particularly important role in supporting the unique metabolic demands of RGCs (Calkins, 2013). Astrocytes show phagocytic activity (for review, see Clarke and Barres, 2013) both in development and in adulthood, with those in the optic nerve head maintaining the extracellular environment by constitutively phagocytosing axons and organelles (Nguyen et al., 2011), including RGC mitochondria (Davis et al., 2014). Because Müller cells are also known for the phagocytic activity (for recent review, see Bejarano-Escobar et al., 2017), both of these classes of retinal (Müller) and optic nerve (astrocyte) glia likely contribute to debris scavenging and microenvironmental homeostasis on an even larger scale than previously appreciated, mimicking to some extent the actions of immune cells that enter the diseased CNS. Failure of these phagocytic roles for glia may promote cell or axon loss over time or inhibit the regenerative capacity of the microenvironment.

In retinal degeneration, like elsewhere in the CNS, glia undergo morphologic changes including stages of hypertrophy and changes in gene expression in a process collectively known as gliosis. Increased expression of the intermediate filament protein, glial filamentous acidic protein (GFAP), is a common hallmark of a late stage of gliosis, but we now appreciate that complex molecular mechanisms likely precede and initiate these profound cellular changes. Gliosis in other parts of the CNS has both adaptive and maladaptive functions. Adaptive functions include regulation and restriction of inflammation and modulation of excitatory activity, whereas maladaptive effects can interfere with the ability of glia to perform their homeostatic functions, making the modulation of

different aspects of gliosis a potential therapeutic target (Sofroniew, 2015). More study is needed about such mechanisms in the retina, optic nerve head, and optic nerve proper.

Despite the abundance and central role of glia in retinal health and disease, there are few diseases known to directly result from glial defects. One possible exception is idiopathic juxtafoveolar retinal telangiectasis [also known as macular telangiectasia type 2 (MacTel-2); Gass and Blodi, 1993]. The histopathology of retinas from MacTel-2 patients has demonstrated loss of Müller cells in affected retinal regions (Powner et al., 2010, 2013); the mechanistic cause and consequence of these changes remains unknown.

Microglia and neuroinflammation

In seeking a better understanding of retinal microenvironments and how to manipulate them, there is perhaps no better place to start than endogenous immune cells. Microglia, the resident immune cells of the CNS, constantly surveil their local microenvironment, extending and retracting their branched processes primarily within the synaptic layers of the retina. In response to changes in the immediate milieu (cytokines, complement, extracellular ions like potassium released from damaged cells), microglia transform into migratory, amoeboid-shaped cells with greater phagocytic capacity, often referred to as an “activated” state. Activated microglia rapidly respond to sites of injury/degeneration where they engulf stressed or dying cells and debris. In reality, there is an entire spectrum of microglial morphologies between ramified and amoeboid microglial states, making morphologic categorization of limited utility when trying to quantify the degree of neuroinflammation, particularly in chronic diseases with on-going slow degeneration and resolution of inflammation in neighboring regions. Histologically it is sometimes also difficult to discern whether apparent engulfment of neurons by microglia is a manifestation of the high rate of phagocytosis that activated microglia typically display, or whether such contacts might sometimes be providing neurotrophic support. Newly developed approaches for single-cell RNA sequencing, transcriptional and epigenomic profiling of rodent and human microglia, such as DropSeq, is revealing molecular signatures that may help identify specific functional states in health and disease (Butovsky et al., 2014; Matcovitch-Natan et al., 2016; Gosselin et al., 2017; Keren-Shaul et al., 2017).

Microglia, like macrophages, can take on either helpful or harmful roles under different conditions, and the signaling mechanisms that mediate these transitions are poorly understood. In healthy CNS tissue, microglia perform homeostatic and physiologic functions that are crucial for CNS development and for regulating neuroplasticity in the adult. For example, ramified microglia prune unneeded synaptic contacts during development (Paolicelli et al., 2011; Schafer et al., 2012) and mediate experience-dependent plasticity in the visual system (Tremblay et al., 2010). In the healthy adult retina, ablation of microglia leads to a reduction in synaptic transmission and ultrastructural inclusions within photoreceptor synap-

tic terminals, revealing an essential role for microglia in synaptic maintenance in the retina (Wang et al., 2016). Conversely, inappropriate activation of microglia mediates loss of synapses in a mouse model of Alzheimer's disease (Hong et al., 2016) and related diseases (Lui et al., 2016; Vasek et al., 2016), reinforcing the view that microglia are homeostatic, microenvironmental biosensors for the CNS.

Interactions between microglia, astroglia, and blood vessels likely help to globally modulate the cellular environment. The Barres Lab (Liddelow et al., 2017) recently showed that when activated, microglia upregulate a number of factors that include TNF- α , IL1 α , and C1q; these molecules in turn cause astrocytes to assume a toxic “A1” state in which they produce other factors, as yet unidentified, that lead to the death of RGCs. Simultaneous deletion of the genes encoding these factors or antibodies binding to their protein product lead to strong neuroprotection of RGCs after injury to the optic nerve. Surprisingly, however, another group recently reported that deletion of microglia with the drug PLX5622 does not alter RGC fate after optic nerve injury (Hilla et al., 2017), suggesting that cells other than microglia might also produce the factors responsible for transforming astrocytes. These findings underscore the importance of increasing our understanding of the cellular interactions involving microglia, other inflammatory cells, and astrocytes after CNS damage.

All neurodegeneration appears to involve microglia (Hanisch and Kettenmann, 2007), which can further escalate the immune response by signaling monocytes to extravasate from retinal vessels and migrate to the site of injury. Once in the retinal environment, monocytes rapidly stop expressing some characteristic monocytic markers, like CCR2, and instead express microglial markers (O’Koren et al., 2016), making it difficult to distinguish the fates of these two cell populations during the course of degeneration. These infiltrative neutrophils and macrophages express a host of trophic and cytotoxic factors, of which the small Ca²⁺-binding protein oncomodulin plays a particularly important role in promoting axon outgrowth (Yin et al., 2006). In age-related macular degeneration (AMD) and photoreceptor degeneration, the accumulation of phagocytes in the subretinal space and surrounding drusen can alter the inhibitory, immunosuppressive nature of the microenvironment, leading to disease progression (for review, see Guillonnet et al., 2017). Until recently, it was thought that there was very little turnover of microglia in the healthy CNS over an organism's lifespan. However, a recent study reported on-going apoptosis and compensatory proliferation (replacement) of microglia in the adult brain (Askew et al., 2017). The differential functions of microglia and monocyte/macrophages in chronic degenerative disease, in age-related susceptibility to disease, and in inflammation following an acute insult is an intense area of research in the CNS in general (Baufeld et al., 2017).

Because microglia play a role in synapse elimination as well as neurodegeneration, they may contribute to synaptic reorganization after transplantation. After optic nerve

injury, the survival of RGCs and/or regeneration of their axons is suppressed by the accumulation of mobile zinc in the terminals of amacrine cells and subsequent exocytotic transfer of this zinc to RGCs (Li et al., 2017). During development, RGCs lose their intrinsic ability for robust axonal growth coincident with the time they receive synaptic inputs from interneurons, and cell culture studies suggest that physical contact between RGCs and amacrine cells, the inhibitory interneurons of the retina, play a role (Goldberg et al., 2002a,b). Thus, it may be worth investigating whether synapse elimination or reorganization via microglia or other phagocytic cells normally occurs after optic nerve damage.

The retinal ECM and the BRB

In the retina, the Müller glial cells are principally responsible for the ECM barriers that construct distinct barriers and microenvironments. Their distal endfeet secrete the collagen-rich basement membrane that constitutes the ILM along the vitreoretinal surface, while their most apical processes envelope the photoreceptor inner segments and form adherens junctions between themselves, forming the external limiting membrane (ELM). Both the ILM and ELM provide structural and diffusional barriers that create microenvironment compartments for the inner and outer retina, respectively (Fig. 1). Within the space between the RPE and ELM, a unique ECM known as the interphotoreceptor matrix (IPM) helps to maintain photoreceptor viability and alignment (Ishikawa et al., 2015). Understanding the distinct composition of these compartments could lead to the control of cell adhesion and motility, affecting both degeneration, repair and regeneration following transplantation. Work in this area is very promising: for example, hybrid scaffolds composed of IPM component and synthetic biopolymers can influence progenitor cell proliferation and promote differentiation toward a photoreceptor-like fate (Baranov et al., 2014).

The molecular details of the matrix comprising the interstitial space in the healthy retina is not well known, though defects in components of the ECM are associated with degeneration and/or functional deficits (for review, see Al-Ubaidi et al., 2013). There may be heterogeneity between the permissiveness of outer and inner retinal matrices, since bipolar cell dendrites can sprout in aging and disease, but RGCs do not. There is also heterogeneity across the surface of the retina, with macular holes healing and filling in, whereas peripheral holes do not. The basis for these differences is not known, but the ECM is likely to be part of the equation.

The retinal blood supply stems from both inner and outer vascular beds: the choriocapillaris, a system of fenestrated capillaries that lies beyond the outer retina and beyond the RPE and Bruch's membrane, and the retinal vessels emanating from the optic nerve head along the inner retinal surface. The outer BRB is primarily Bruch's membrane and the tight-junctions between the RPE cells, which prevent the unregulated passage of cells, proteins and ions into the subretinal space. The inner BRB relies on the tight junctions between the vascular endothelial cells themselves, which are affected by

Table 1. Specific recommendations from workshop participants

-
- Development of tools for labeling/manipulating astrocytes in the living retina
 - Gene expression profiling of glial cell types during gliosis
 - A better understand of Müller cell roles in creating and maintaining ECM and retinal health
 - Examination of the transcription factors/signaling mechanisms that control microglial state changes
 - Methods to label resident and systemic immune cells acutely in order to examine morphology during disease (ideally suitable for use in humans)
 - Further understanding of microglia signaling mechanisms during aging and disease
 - Methods to make transplanted cells less adherent to CSPGs at scarred sites
 - Further development of ECM scaffolds to promote regeneration or integration of transplanted cells
 - The ability to manipulate the blood-retina barrier in a region-specific manner for targeted delivery of therapeutic agents
-

inflammation, glycation, and edema (Klaassen et al., 2013). Like elsewhere in the CNS, blood flow and leakage across the BRB is regulated by inner retinal astrocytes, vascular pericytes and perivascular macrophages, and BRB breakdown is associated with many chronic retinal diseases. The molecular mechanisms that maintain a healthy barrier and are affected in disease appear to be somewhat different from those for the blood-brain barrier and remain relatively understudied (Díaz-Coránguez et al., 2017). Two of the most prevalent diseases of the retina, wet AMD and diabetic retinopathy, lead to breaches of the outer and inner BRB, respectively. Thus, the motivation for understanding how the BRB is maintained and how it might be restored in disease is very high.

Next steps

Many gaps and barriers currently limit our understanding of retinal glia, microglia and inflammation, and the ECM and BRB. First, the role of glia in regulating the retinal microenvironment in both health and disease needs additional study. For example, unlike glia elsewhere in the CNS, there is very little known about the roles of Müller cells at retinal synapses, of astrocytes signaling with RGC axons, and of the normal rate of glial phagocytosis or synaptic maintenance in healthy retina. Are there subtypes or "states" of reactive astrocytes in the retina that exert neurotoxic or neuroprotective effects like in the brain (Liddelow et al., 2017)? Do retinal astrocytes control lymphatic perfusion of the interstitial space like in the brain (Jessen et al., 2015)? To what degree can neuroregeneration be augmented by targeting both astrocyte and microglial activation states?

To tackle these questions as a field, additional tools and resources are needed (Table 1). Although there are several good markers for labeling Müller cells in the retina, including a custom AAV called Shh10 (Klimczak et al., 2009) that drives serotype-specific Müller cell transduction, there is a need for consistent analogous tools for labeling or manipulating astrocytes exclusively in the liv-

ing retina or optic nerve head. Gene expression profiling of both glial cell types before and during degeneration will help to define the sequence of signaling events that precede frank gliosis, and which may contribute to early loss of neuronal function. Such expression profiling may help to identify drug targets that could in turn be harnessed to control glial function throughout disease progression, both by boosting glial-based metabolic/neurotrophic support and by delaying or reversing gliosis itself. Finally, a turn away from typical rodent-based model systems could dramatically change our understanding of Müller cell function, since Müller cells in lower vertebrates readily regenerate or transform to a progenitor-like state (Goldman, 2014). Emerging research indicates a similar, though far more limited, capacity in rodents, which we may learn to harness by comparisons with lower vertebrates. Understanding this regenerative power is the first step to harnessing its potential for control of the microenvironment and improving transplantation efficiency.

With regards to microglia, because they are motile and dynamic in both form and function, they offer great potential for cell transplantation and neuroregeneration fields. Since microglia are essential for reinforcing and sustaining proper synaptic connections in healthy tissue, they perhaps could likewise be programmed to prune away the stressed or damaged regions of the retina. Achieving this will require a better understanding of the transcription factors or signaling mechanisms that control microglial state changes. Understanding these signaling networks could also be harnessed to develop therapeutics that could boost microglial support functions and delay phagocytic transformation. Developing such reagents is challenging, in part because of the very nature of microglia's sensitivity to their environment: microglia steadily transform *ex vivo* into an activated phenotype. To study microglia *in vivo*, high-resolution retinal imaging methods like adaptive-optics scanning laser ophthalmoscopy (AO-SLO) can resolve microglial morphology and branch dynamics across different retinal layers in *Cx3CR1^{gfp/+}* mice (Zawadzki et al., 2015). While this genetic approach in mice is experimentally convenient, it would be better to label resident and systemic immune cells acutely, which could then be applied to any species, including human subjects in any state of retinal disease. Unlike nearly all other cell types in the retina, to date there are no known viral serotypes or nanoparticle tagging methods that reliably reveal microglia within their native environment. A better understanding of the signaling mechanisms of microglia, and how they change in aging and disease, might allow us to program resident cells to migrate to and deliver support as needed.

Our understanding of the ECM in the retina is limited. As in other tissues, deposition of chondroitin sulfate proteoglycans (CSPGs) generally create sticky barriers through which cells cannot readily migrate or axons readily penetrate. In injury and disease, gliosis leads to disruption of ECM homeostasis and deposition of CSPGs and other inhibitory glycans (neurocan, aggrecan, versican), which will need to be overcome for transplantation

therapies to efficiently lead to cell migration, integration and synaptogenesis (Lau et al., 2013). Currently, best strategies for overcoming CSPGs is to simply digest with matrix metalloproteinase-2 (MMP-2), though this non-specific destruction might be problematic because some matrix scarring can prevent the spread of damage. A better alternative would be to make transplanted cells less adherent to CSPGs, making them "blind" to the scarred site.

Studies which define the properties of the interstitial/ECM environment over the course of development and aging could provide important clues to explain why adult tissue is so anti-regenerative. The retina virtually explodes with proliferation during development, with many more neurons being born than are ultimately needed, creating a wave of apoptosis and synaptogenesis that likely drives up the phagocytic activity of glia and microglia alike. Can we exploit the ECM/microenvironment of the developing optic projection (nerve and retina) to better understand why it is not permissive in adulthood? Can we harness the latent regenerative capacity of Müller cells (Sifuentes et al., 2016; Jorstad et al., 2017) or re-engage a development-like state by tweaking the ECM where it is needed during degeneration? Can we engineer specific ECM scaffolds to promote regeneration or integration of transplanted cells or to allow cell and process motility while preserving the benefits of CSPG barriers to isolate damaged areas? How can a healthy BRB be best supported throughout aging and disease? The ability to manipulate the ECM and the BRB in a region-specific manner could be key to modulate glial reactivity and neuroinflammation in disease, and to deliver therapeutic agents from the bloodstream directly to regions of injury and degeneration.

Summary

To optimize the cellular environment for neuroregeneration and cell transplantation, much more basic knowledge is needed about individual retinal niches, the composition of the interstitial spaces within the retina (e.g., ECM), the optic nerve head, and the optic nerve and how these spaces are normally maintained by the BRB, by glia, and by microglia. Comparisons of these niches through the course of development and aging will inform us of candidate mechanisms to promote growth-friendly environments for regeneration. Each of the fields of study individually have specific needs and strengths (Table 1), though clearly there is a need for collaboration between experts within different niches to accelerate discovery.

Several overarching principles readily emerged from the group's consideration of the state of the field *in toto*. First, this general area woefully lacks the same depth of basic mechanistic understanding enjoyed by some other fields. Investigators should consider more basic science research and fewer disease-based studies. The rationale for this is that normal function of glia, microglia and the neurovascular unit is likely to be similar across species, whereas individual diseases may produce uniquely specific changes that render common mechanisms difficult to

infer. Second, each retinal niche suffers from a paucity of descriptive or cataloged definitions. For example, each niche needs to address issues of cell-type specificity and heterogeneity, and how these populations change during development, aging and disease, in both animal models and humans. DropSeq and other high-throughput methods that are often beyond the means of a single lab or insufficiently hypothesis-driven for a typical individual investigator award are nonetheless vital for identifying the building blocks on which new hypotheses for mechanism can be generated.

Finally, such building blocks of knowledge would presumably lead to the development of new tools which are vital for monitoring and controlling cellular environments. New and better animal models are needed to bridge our understanding from mice to humans. Centralized, managed database repositories and a nationalized system for donor tissue banks would facilitate resource and data sharing in a manner that would ultimately save time and money and promote collaboration. As one participant pointed out, the field of neuroregeneration needs the “optogenetics equivalent” for tweaking glia and microglia to promote cell survival, tissue remodeling, and synaptogenesis as needed. Through cross-fertilization of fields, collaborative approaches, and investment of resources in basic discovery science, manipulation of the cellular environments to promote regeneration can soon be a reality.

References

- Adams KL, Gallo V (2018) The diversity and disparity of the glial scar. *Nat Neurosci* 21:9–15. [CrossRef](#) [Medline](#)
- Al-Ubaidi MR, Naash MI, Conley SM (2013) A perspective on the role of the extracellular matrix in progressive retinal degenerative disorders. *Invest Ophthalmol Vis Sci* 54:8119–8124. [CrossRef](#)
- Askew K, Li K, Olmos-Alonso A, Garcia-Moreno F, Liang Y, Richardson P, Tipton T, Chapman MA, Riecken K, Beccari S, Sierra A, Molnár Z, Cragg MS, Garaschuk O, Perry VH, Gomez-Nicola D (2017) Coupled proliferation and apoptosis maintain the rapid turnover of microglia in the adult brain. *Cell Rep* 18:391–405. [CrossRef](#)
- Baranov P, Michaelson A, Kundu J, Carrier RL, Young M (2014) Interphotoreceptor matrix-poly(ϵ -caprolactone) composite scaffolds for human photoreceptor differentiation. *J Tissue Eng* 5:2041731414554139. [CrossRef](#)
- Baufeld C, O’Loughlin E, Calcagno N, Madore C, Butovsky O (2017) Differential contribution of microglia and monocytes in neurodegenerative diseases. *J Neural Transm (Vienna)*. [CrossRef](#)
- Bejarano-Escobar R, Sanchez-Calderon H, Otero-Arenas J, Martin-Partido G, Francisco-Morcillo J (2017) Muller glia and phagocytosis of cell debris in retinal tissue. *J Anat* 231:471–483.
- Bringmann A, Pannicke T, Grosche J, Francke M, Wiedemann P, Skatchkov SN, Osborne NN, Reichenbach A (2006) Müller cells in the healthy and diseased retina. *Prog Retin Eye Res* 25:397–424. [CrossRef](#)
- Büssow H (1980) The astrocytes in the retina and optic nerve head of mammals: a special glia for the ganglion cell axons. *Cell Tissue Res* 206:367–378. [CrossRef](#)
- Butovsky O, Jedrychowski MP, Moore CS, Cialic R, Lanser AJ, Gabrieli G, Koeglperger T, Dake B, Wu PM, Doykan CE, Fanek Z, Liu L, Chen Z, Rothstein JD, Ransohoff RM, Gygi SP, Antel JP, Weiner HL (2014) Identification of a unique TGF- β -dependent molecular and functional signature in microglia. *Nat Neurosci* 17:131–143. [CrossRef](#)
- Calkins DJ (2013) Age-related changes in the visual pathways: blame it on the axon. *Invest Ophthalmol Vis Sci* 54:ORSF37–ORSF41. [CrossRef](#) [Medline](#)
- Choi HJ, Sun D, Jakobs TC (2015) Astrocytes in the optic nerve head express putative mechanosensitive channels. *Mol Vis* 21:749–766.
- Clarke LE, Barres BA (2013) Emerging roles of astrocytes in neural circuit development. *Nat Rev Neurosci* 14:311–321. [CrossRef](#) [Medline](#)
- Davis CH, Kim KY, Bushong EA, Mills EA, Boassa D, Shih T, Kinebuchi M, Phan S, Zhou Y, Bihlmeyer NA, Nguyen JV, Jin Y, Ellisman MH, Marsh-Armstrong N (2014) Transcellular degradation of axonal mitochondria. *Proc Natl Acad Sci USA* 111:9633–9638. [CrossRef](#)
- Díaz-Coránguez M, Ramos C, Antonetti DA (2017) The inner blood-retinal barrier: cellular basis and development. *Vis Res* 139:123–137. [CrossRef](#)
- Eiraku M, Takata N, Ishibashi H, Kawada M, Sakakura E, Okuda S, Sekiguchi K, Adachi T, Sasai Y (2011) Self-organizing optic-cup morphogenesis in three-dimensional culture. *Nature* 472:51–56. [CrossRef](#)
- Fischer AJ, Bongini R (2010) Turning Muller glia into neural progenitors in the retina. *Mol Neurobiol* 42:199–209. [CrossRef](#)
- Gass JD, Blodi BA (1993) Idiopathic juxtafoveolar retinal telangiectasis. Update of classification and follow-up study. *Ophthalmology* 100:1536–1546. [CrossRef](#)
- Giannoni P, Badaut J, Dargazanli C, De Maudave AF, Klement W, Costalat V, Marchi N (2018) The pericyte-glia interface at the blood-brain barrier. *Clin Sci (Lond)* 132:361–374. [CrossRef](#)
- Goldberg JL, Espinosa JS, Xu Y, Davidson N, Kovacs GT, Barres BA (2002a) Retinal ganglion cells do not extend axons by default: promotion by neurotrophic signaling and electrical activity. *Neuron* 33:689–702. [CrossRef](#)
- Goldberg JL, Klassen MP, Hua Y, Barres BA (2002b) Amacrine-signaled loss of intrinsic axon growth ability by retinal ganglion cells. *Science* 296:1860–1864. [CrossRef](#)
- Goldman D (2014) Müller glial cell reprogramming and retina regeneration. *Nat Rev Neurosci* 15:431–442. [CrossRef](#) [Medline](#)
- Gosselin D, Skola D, Coufal NG, Holtman IR, Schlachetzki JCM, Sajti E, Jaeger BN, O’Connor C, Fitzpatrick C, Pasillas MP, Pena M, Adair A, Gonda DD, Levy ML, Ransohoff RM, Gage FH, Glass CK (2017) An environment-dependent transcriptional network specifies human microglia identity. *Science* 356. [CrossRef](#)
- Guillonneau X, Eandi CM, Paques M, Sahel JA, Sapiéha P, Sennlaub F (2017) On phagocytes and macular degeneration. *Prog Retin Eye Res* 61:98–128.
- Hanisch UK, Kettenmann H (2007) Microglia: active sensor and versatile effector cells in the normal and pathologic brain. *Nat Neurosci* 10:1387–1394. [CrossRef](#)
- Hilla AM, Diekmann H, Fischer D (2017) Microglia are irrelevant for neuronal degeneration and axon regeneration after acute injury. *J Neurosci* 37:6113–6124. [CrossRef](#)
- Hong S, Beja-Glasser VF, Nfonoyim BM, Frouin A, Li S, Ramakrishnan S, Merry KM, Shi Q, Rosenthal A, Barres BA, Lemere CA, Selkoe DJ, Stevens B (2016) Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science* 352:712–716. [CrossRef](#)
- Hurley JB, Lindsay KJ, Du J (2015) Glucose, lactate, and shuttling of metabolites in vertebrate retinas. *J Neurosci Res* 93:1079–1092. [CrossRef](#)
- Ishikawa M, Sawada Y, Yoshitomi T (2015) Structure and function of the interphotoreceptor matrix surrounding retinal photoreceptor cells. *Exp Eye Res* 133:3–18. [CrossRef](#)
- Jessen NA, Munk ASF, Lundgaard I, Nedergaard M (2015) The glymphatic system: a beginner’s guide. *Neurochem Res* 40:2583–2599. [CrossRef](#) [Medline](#)
- Jo AO, Ryskamp DA, Phuong TT, Verkman AS, Yarishkin O, MacAulay N, Krizaj D (2015) TRPV4 and AQP4 channels synergistically regulate cell volume and calcium homeostasis in retinal Muller glia. *J Neurosci* 35:13525–13537. [CrossRef](#)

- Jorstad NL, Wilken MS, Grimes WN, Wohl SG, VandenBosch LS, Yoshimatsu T, Wong RO, Rieke F, Reh TA (2017) Stimulation of functional neuronal regeneration from Müller glia in adult mice. *Nature* 548:103–107. [CrossRef](#)
- Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Sternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, Itzkovitz S, Colonna M, Schwartz M, Amit I (2017) A unique microglia type associated with restricting development of Alzheimer's disease. *Cell* 169:1276–1290.e1217. [CrossRef](#)
- Klaassen I, Van Noorden CJ, Schlingemann RO (2013) Molecular basis of the inner blood-retinal barrier and its breakdown in diabetic macular edema and other pathological conditions. *Prog Retin Eye Res* 34:19–48. [CrossRef](#)
- Klimczak RR, Koerber JT, Dalkara D, Flannery JG, Schaffer DV (2009) A novel adeno-associated viral variant for efficient and selective intravitreal transduction of rat Müller cells. *PLoS One* 4:e7467. [CrossRef](#)
- Lau LW, Cua R, Keough MB, Haylock-Jacobs S, Yong VW (2013) Pathophysiology of the brain extracellular matrix: a new target for remyelination. *Nat Rev Neurosci* 14:722–729. [CrossRef](#) [Medline](#)
- Lenkowski JR, Raymond PA (2014) Müller glia: stem cells for generation and regeneration of retinal neurons in teleost fish. *Prog Retin Eye Res* 40:94–123. [CrossRef](#)
- Li Y, Anderegg L, Yuki K, Omura K, Yin Y, Gilbert HY, Erdogan B, Asdourian MS, Shrock C, de Lima S, Apfel UP, Zhuo Y, Hershfinkel M, Lippard SJ, Rosenberg PA, Benowitz L (2017) Mobile zinc increases rapidly in the retina after optic nerve injury and regulates ganglion cell survival and optic nerve regeneration. *Proc Natl Acad Sci USA* 114:E209–E218. [CrossRef](#)
- Liddel SA, Gattenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, Bennett ML, Münch AE, Chung WS, Peterson TC, Wilton DK, Frouin A, Napier BA, Panicker N, Kumar M, Buckwalter MS, Rowitch DH, Dawson VL, Dawson TM, Stevens B, et al. (2017) Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541:481–487. [CrossRef](#)
- Lui H, Zhang J, Makinson SR, Cahill MK, Kelley KW, Huang HY, Shang Y, Oldham MC, Martens LH, Gao F, Coppola G, Sloan SA, Hsieh CL, Kim CC, Bigio EH, Weintraub S, Mesulam MM, Rademakers R, Mackenzie IR, Seeley WW, et al. (2016) Progranulin deficiency promotes circuit-specific synaptic pruning by microglia via complement activation. *Cell* 165:921–935. [CrossRef](#)
- MacVicar BA, Newman EA (2015) Astrocyte regulation of blood flow in the brain. *Cold Spring Harb Perspect Biol* 7. [CrossRef](#)
- Matcovitch-Natan O, Winter DR, Giladi A, Vargas Aguilar S, Spinrad A, Sarrazin S, Ben-Yehuda H, David E, Zelada Gonzalez F, Perrin P, Keren-Shaul H, Gurly M, Lara-Astaiso D, Thaiss CA, Cohen M, Bahar Halpern K, Baruch K, Deczkowska A, Lorenzo-Vivas E, Itzkovitz S, et al. (2016) Microglia development follows a stepwise program to regulate brain homeostasis. *Science* 353:aad8670. [CrossRef](#)
- Meyer JS, Shearer RL, Capowski EE, Wright LS, Wallace KA, McMillan EL, Zhang SC, Gamm DM (2009) Modeling early retinal development with human embryonic and induced pluripotent stem cells. *Proc Natl Acad Sci USA* 106:16698–16703. [CrossRef](#)
- Meyer JS, Howden SE, Wallace KA, Verhoeven AD, Wright LS, Capowski EE, Pinilla I, Martin JM, Tian S, Stewart R, Pattnaik B, Thomson JA, Gamm DM (2011) Optic vesicle-like structures derived from human pluripotent stem cells facilitate a customized approach to retinal disease treatment. *Stem Cells* 29:1206–1218. [CrossRef](#)
- Newman EA (2015) Glial cell regulation of neuronal activity and blood flow in the retina by release of gliotransmitters. *Philos Trans R Soc Lond B Biol Sci* 370. [CrossRef](#)
- Nguyen JV, Soto I, Kim KY, Bushong EA, Oglesby E, Valiente-Soriano FJ, Yang Z, Davis CH, Bedont JL, Son JL, Wei JO, Buchman VL, Zack DJ, Vidal-Sanz M, Ellisman MH, Marsh-Armstrong N (2011) Myelination transition zone astrocytes are constitutively phagocytic and have synuclein dependent reactivity in glaucoma. *Proc Natl Acad Sci USA* 108:1176–1181. [CrossRef](#)
- O'Koren EG, Mathew R, Saban DR (2016) Fate mapping reveals that microglia and recruited monocyte-derived macrophages are definitively distinguishable by phenotype in the retina. *Sci Rep* 6:20636. [CrossRef](#)
- Paolicelli RC, Bolas G, Pagani F, Maggi L, Scianni M, Panzanelli P, Giustetto M, Ferreira TA, Guiducci E, Dumas L, Ragozzino D, Gross CT (2011) Synaptic pruning by microglia is necessary for normal brain development. *Science* 333:1456–1458. [CrossRef](#)
- Pownner MB, Gillies MC, Tretiach M, Scott A, Guymer RH, Hageman GS, Frutiger M (2010) Perifoveal Müller cell depletion in a case of macular telangiectasia type 2. *Ophthalmology* 117:2407–2416. [CrossRef](#)
- Pownner MB, Gillies MC, Zhu M, Vevis K, Hunyor AP, Frutiger M (2013) Loss of Müller's cells and photoreceptors in macular telangiectasia type 2. *Ophthalmology* 120:2344–2352. [CrossRef](#)
- Reichman S, Slembrouck A, Gagliardi G, Chaffiol A, Terray A, Nanteau C, Potey A, Belle M, Rabesandratana O, Duebel J, Orioux G, Nandrot EF, Sahel JA, Goureau O, et al. (2017) Generation of storable retinal organoids and retinal pigmented epithelium from adherent human iPS cells in xeno-free and feeder-free conditions. *Stem Cells* 35:1176–1188. [CrossRef](#)
- Ryskamp DA, Jo AO, Frye AM, Vazquez-Chona F, MacAulay N, Thoreson WB, Krizaj D (2014) Swelling and eicosanoid metabolites differentially gate TRPV4 channels in retinal neurons and glia. *J Neurosci* 34:15689–15700. [CrossRef](#)
- Sarthy V, Ripps H (2001) The retinal Müller cell: structure and function. New York: Kluwer Academic.
- Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, Yamasaki R, Ransohoff RM, Greenberg ME, Barres BA, Stevens B (2012) Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron* 74:691–705. [CrossRef](#)
- Shirai H, Mandai M, Matsushita K, Kuwahara A, Yonemura S, Nakano T, Assawachananont J, Kimura T, Saito K, Terasaki H, Eiraku M, Sasai Y, Takahashi M (2016) Transplantation of human embryonic stem cell-derived retinal tissue in two primate models of retinal degeneration. *Proc Natl Acad Sci USA* 113:E81–E90. [CrossRef](#)
- Sifuentes CJ, Kim JW, Swaroop A, Raymond PA (2016) Rapid, dynamic activation of Müller glial stem cell responses in zebrafish. *Invest Ophthalmol Vis Sci* 57:5148–5160. [CrossRef](#)
- Sofroniew MV (2015) Astrocyte barriers to neurotoxic inflammation. *Nat Rev Neurosci* 16:249–263. [CrossRef](#) [Medline](#)
- Tremblay ME, Lowery RL, Majewska AK (2010) Microglial interactions with synapses are modulated by visual experience. *PLoS Biol* 8:e1000527. [CrossRef](#) [Medline](#)
- Vasek MJ, Garber C, Dorsey D, Durrant DM, Bollman B, Soung A, Yu J, Perez-Torres C, Frouin A, Wilton DK, Funk K, DeMasters BK, Jiang X, Bowen JR, Mennerick S, Robinson JK, Garbow JR, Tyler KL, Suthar MS, Schmidt RE, et al. (2016) A complement-microglial axis drives synapse loss during virus-induced memory impairment. *Nature* 534:538–543. [CrossRef](#)
- Wahlin KJ, Maruotti JA, Sripathi SR, Ball J, Angueyra JM, Kim C, Grebe R, Li W, Jones BW, Zack DJ (2017) Photoreceptor outer segment-like structures in long-term 3D retinas from human pluripotent stem cells. *Sci Rep* 7:766. [CrossRef](#)
- Wang X, Zhao L, Zhang J, Fariss RN, Ma W, Kretschmer F, Wang M, Qian HH, Badea TC, Diamond JS, Gan WB, Roger JE, Wong WT (2016) Requirement for microglia for the maintenance of synaptic function and integrity in the mature retina. *J Neurosci* 36:2827–2842. [CrossRef](#)
- Wiley LA, Burnight ER, DeLuca AP, Anfinson KR, Cranston CM, Kaalberg EE, Penticoff JA, Affatigato LM, Mullins RF, Stone EM, Tucker BA (2016) cGMP production of patient-specific iPSCs and photoreceptor precursor cells to treat retinal degenerative blindness. *Sci Rep* 6:30742. [CrossRef](#)
- Xue Y, Shen SQ, Jui J, Rupp AC, Byrne LC, Hattar S, Flannery JG, Corbo JC, Kefalov VJ (2015) CRALBP supports the mammalian retinal visual cycle and cone vision. *J Clin Invest* 125:727–738. [CrossRef](#) [Medline](#)

- Yin Y, Henzl MT, Lorber B, Nakazawa T, Thomas TT, Jiang F, Langer R, Benowitz LI (2006) Oncomodulin is a macrophage-derived signal for axon regeneration in retinal ganglion cells. *Nat Neurosci* 9:843–852. [CrossRef](#)
- Zawadzki RJ, Zhang P, Zam A, Miller EB, Goswami M, Wang X, Jonnal RS, Lee SH, Kim DY, Flannery JG, Werner JS, Burns ME, Pugh EN Jr (2015) Adaptive-optics SLO imaging combined with widefield OCT and SLO enables precise 3D localization of fluorescent cells in the mouse retina. *Biomed Opt Express* 6:2191–2210. [CrossRef](#)
- Zhong X, Gutierrez C, Xue T, Hampton C, Vergara MN, Cao LH, Peters A, Park TS, Zambidis ET, Meyer JS, Gamm DM, Yau KW, Canto-Soler MV (2014) Generation of three-dimensional retinal tissue with functional photoreceptors from human iPSCs. *Nat Commun* 5:4047. [CrossRef](#)